



Examination of microbial and chemical properties of chicken fillet treated with *Saturejarechingeri* essential oil, sodium dodecyl sulfate and citric acid during refrigerated storage

Siavash Maktabi^{*1}, ZeynabSafdari², Mehdi PoormehdiBroujeni¹

- 1- Department of Food Hygiene, Faculty of Veterinary Medicine, Shahid Chamran University of Ahvaz, Ahvaz, Iran
- 2- Graduated from Faculty of Veterinary Medicine, Shahid Chamran University of Ahvaz, Iran

ARTICLE INFO

ABSTRACT

Article History:

Received:2022/10/11

Accepted: 2023/2/20

Keywords:

Chicken meat;
Chicken fillet;
Saturejarechingeri essential oil;
Sodium dodecyl sulfate;
Citric acid;
Shelf life

DOI: 10.22034/FSCT.20.140.1

DOR:20.1001.1.20088787.1402.20.140.1.7

*Corresponding Author E-Mail:
s.maktabi@scu.ac.ir

The effects of *Saturejarechingeri* essential oil (*Sr*-EO), sodium dodecyl sulfate (SDS) and citric acid on increasing the shelf life of chicken fillets during refrigerated storage were investigated. The treatments included 28 chicken fillets divided into four groups, with three replicates each. Each group was immersed in a solution containing normal saline (control), 0.5% *Sr*-EO, 0.5% *Sr*-EO + 0.5% SDS, or 0.5% *Sr*-EO + 0.5% SDS + 0.1% citric acid for 15 min and refrigerated for 15 days. Bacterial quality (mesophilic and psychrophilic total counts), chemical analysis (pH, TBA, and TVN), and sensory evaluation were performed on days 0, 3, 6, 9, 12, and 15. The bacterial load of mesophilic and psychrophilic bacteria increased over time for all four groups, but the number of bacteria in *Sr*-EO + SDS + citric acid group for mesophilic microbial index and in *Sr*-EO + SDS group for psychrophilic microbial index was less than that in other samples. At the end of storage time, the highest TVN value was observed for control group (41.06 ± 8.29 g / 100 g) and the lowest value (33 mg / 100 g) was found for *Sr*-EO+SDS +citric acid group followed by *Sr*-EO+SDS and *Sr*-EO groups. There were no significant differences in the TBA value and pH between the treatment and control samples. The control group showed undesirable and unacceptable sensory properties on day 6 whereas the sensory factors of *Sr*-EO and SDS+*Sr*-EO groups were within the optimal range of consumer acceptance until days 9 and 6, respectively. The results showed that compared to controls, the SDS+*Sr*-EO solution was more effective in extending the shelf life of refrigerated chicken fillets to four days. This type of combined treatment could be a useful way to increase the refrigerated storage time of chicken meat.

1- Introduction

Chicken is a highly perishable food that will deteriorates within 4-10 days even when it is refrigerated [1]. Fresh meat products are usually marketed at refrigerated temperature (2-5°C). Lipid oxidation and microbial growth can occur during refrigerated storage. Chicken meat spoilage can cause huge economic losses to producers; therefore new methods need to be developed to extend the shelf life and improve the overall quality of meat [2]. To retard or minimize the oxidative spoilage of foods, effective synthetic antioxidants are added to the products; however, owing to their potential carcinogenesis [3]. and the lack of consumer acceptance, natural antioxidants have recently attracted much attention; So, there is a growing interest in identifying new and natural antioxidants that can be used in place of synthetic compounds [4].

Saturejarechingeri belonging to the *Lamiaceae* family, is one of an exclusive savory species in southwestern Iran [5]. *S.rechingeri* is commercially important (medicinal, food, and cosmetic applications) due of its high carvacrol content in the essential oils and free phenolic acids [6]. In traditional medicine, it is used as a sedative, disinfectant, decoction, and spice. In recent years, extensive studies have been conducted on the biological and medicinal effects of this plant including its antioxidant, antimicrobial, anti-diabetic, analgesic and anti-inflammatory properties and interesting results have been obtained. Its antimicrobial effects have been studied in vitro and demonstrated by many researchers. Carvacrol and thymol have been reported to play key roles in the antimicrobial activity of *S. rechingeri* essential oil (*Sr-EO*). Thymol and carvacrol are also found in other plants, such as thyme, but their high proportion in *S.rechingeri* is responsible

for their characteristic antimicrobial effects [7,8].

Sodium dodecyl sulfate (SDS) is a potent alkaline sulfate microbicide and a highly effective anionic surfactant that kills bacteria by denaturing cell surface proteins. SDS is also a common ingredient used in cosmetics and detergents and is used in the laboratory as a protein solvent. It is a safe and healthy substance, with no adverse effects on human health [9]. In recent years, the use of SDS as a disinfectant in the food and equipment industry has attracted increasing attention. The effect of SDS, individually or in combination with other substances on reducing bacterial contamination has been studied in beef [10], chicken breast meat, and blueberry [11,12]. The use of citric acid at standard concentrations as a flavoring in food is common and harmless. The use of citric acid along with other disinfectants has a synergistic effect on reducing microbial count [13, 14].

To date the effect of *Sr-EO* individually or in combination with SDS and citric acid, on the shelf life of chicken meat has not been studied. Therefore, in the present study, we investigated the effects of *Sr-EO*, SDS, and citric acid on the shelf life of chicken meat.

2. Materials & methods

2. 1.Extraction of essential oil and analysis of its compounds

The leaves of *S.rechingeri* were collected from Dehloran region located in Ilam province, separated from the stem, dried in the shade, and then crushed in a mill. The essential oil was extracted using the water distillation method and a Clevenger apparatus. The essential oil yield was 3.41% (3.41 ml essential oil per 100 g of dried plants). The prepared essential oil was injected into a GC/MS instrument (Agilent 5977B, USA) to characterize its constituents. The capillary column was HP-5MS (5% phenyl methyl

silicone and 95% dimethylpolysiloxane), (length: 30m; internal diameter: 0.25mm and 0.25 μ m film thickness). The column temperature program was set as follows: the oven temperature was maintained at 60 °C for 1 min, changed from 60°C to 250 °C at 5 °C/min, and maintained at 250 °C for 2 min. The Injection volume was 0.2 μ L and the temperature of the injector was 250°C. The carrier gas was helium with a purity of 99.99%, constant flow rate 1.1ml per min, and split ratio of 1:100. . The procedure was operated in electron impact mode at 70 eV. GC-MS analysis was performed in triplicate.

2. 2.Preparation and treatments of samples

Fresh chicken fillets were purchased from Ahwaz retail market on the day of slaughter. 28 portions of fillets weighing 100-120 g were separated and manually prepared for treatment. They were washed with tap water and placed on a UV-sterilized stainless steel colander to remove excess water. The colander was then placed in laminar hood for 5 min. The fillets were divided into four groups and placed separately in UV-sterilized plastic containers, and 200 mL of each treatment solution was transferred to each container (Table 1). Chicken fillets were immersed in the solution for 15 min at room temperature.

Table 1: Different treatment solutions

Treatment 1	Normal saline as control
Treatment 2	0.5% <i>Sr</i> -EO
Treatment 3	0.5% <i>Sr</i> -EO + 0.5% SDS
Treatment 4	0.5% <i>Sr</i> -EO + 0.5% SDS +0.1% citric acid

After 15 min, the fillets were placed in sterile UV-sterilized polyethylene bags. A total of 28 fillets (seven pieces per treatment) were used each time. All samples were refrigerated at 7°C for 15 d. Sampling was performed on days 0, 3, 6, 9, 12, and 15, and microbial, chemical, and

sensory evaluations were performed. All experiments were performed in triplicates.

2.3.Microbiological analysis

Chicken fillets (10g) were aseptically transferred into sterile bags containing 90 ml of normal saline and homogenized in a stomacher (Lab blender 400, France) for 1 min. Serial dilutions (1:10) were prepared and each diluted sample (0.1 ml) was cultured on plate count agar. Plates were incubated for 48 h at 37°C and for 10 day at 7°C to count mesophilic and psychrophilic bacteria, respectively [15,16]. The results were expressed as log colony forming units per gram (log CFU/g) [17].

2. 4. Determination of thiobarbituric acid reactive substance (TBARS)

Following Xiong et al. (2015) with some modifications, 5 g of the sample was homogenized in 50 ml of sterile distilled water. Five gram of Trichloroacetic acid powder was added and mixed rapidly to prevent lumping. The mixture was then filtered and trichloroacetic acid solution (10%) was added until the volume reached 50 ml. Then, 3 ml of the filtered solution and 3 ml of 0.2 M thiobarbituric acid reagent were mixed in a glass tube and placed in an oven at 95 °C for 45 min. After cooling, the absorbance of the samples was measured at 532 nm and the amount of malondialdehyde (mg/kg of fillet) was calculated using the following equation [18]:

$$\text{TBRS (mg MDA/kg)} = \text{Absorbance rate} \times 7.8$$

2. 5. Measurement of total volatile nitrogen

Volatile nitrogen content was achieved by Kjeldal apparatus. The chicken samples (5 g) and distilled water (60 mL) were mixed and transferred into a tube containing 1 g of magnesium oxide (MgO). The distillate was absorbed using 40 ml aqueous solution of 2% boric acid containing 0.5

ml of tashiro's indicator solution. The boric acid solution was titrated using sulfuric acid (0.1 N) solution, and the amount of acid consumed was recorded. The total volatile nitrogen was calculated in mg per 100 g of chicken meat using the following equation[19, 20];

$$\text{TVN (mg/100 g)} = (\text{consumed acid (ml)} / \text{sample-consumed acid (ml) of control} \times 1/4 \times 100) / (\text{weight of sample (g)})$$

2. 6. Measurement of pH

Chicken samples(10g)were homogenized in 90 ml of distilled water. The pH valuesof the samples were measured using a digital pH meter[21,22].

2. 7. Changes in sensory properties

Raw chicken fillets were evaluated in terms of color, odor, texture,and overall acceptance by a panel offive people (five women, 25-35 years old). Sensory evaluation was performed using a 5-point hedonic scoring system including color (5, no color changes; 1, severe color changes), odor (5, highly desirable; 1,unacceptable), texture (5, firm; 1, very soft), and total acceptance (5, highly desirable, 1, highly unacceptable). The acceptance score for human consumption of chicken fillets was 4[23, 24].

2. 8. Statistical analysis

Data were analyzed descriptively and analytically using SPSS version 16. Quantitative data analysis (TBA, TVN, PH and logarithm of mesophilic and psychrophilic bacteria) were performed by variance analysis with repeated measurements and LSD supplementary tests. Qualitative data analysis (odor, color, texture, and total acceptance) was performed using Friedman and Kruskal-Wallis tests. Statistical significance was set at $p \leq 0.05$.

3. Results

3.1. Chemical compositions of the Sr-EO

The constituents of Sr-EO were analyzed by GC/MS are presented in Table 2. The main compounds were carvacrol (86.91%), gamma-terpinene(2.84%), *p*-cymene (2.08%) and carvacryl acetate (1.29%).

Table.2. Chemical compositions of the Sr-EO

N	Compound	%	RI	RT
1	Alpha-tujene	0.36	924	5.011
2	Alpha-pinene	0.2	932	5.165
3	Beta-myrcene	0.7	988	6.365
4	Phellandrene	0.12	1002	6.711
5	Alpha-terpinene	0.73	1014	7.006
6	P-cymene	2.08	1020	7.199
7	Gamma.terpinen	2.84	1054	8.059
8	Cis-sabinenehydrat	0.19	1065	8.283
9	Alpha-terpinolene	0.1	1086	8.828
10	Linalol	0.55	1095	9.098
11	Borneol	0.15	1165	10.913
12	Terpinene-4-ol	0.81	1174	11.221
13	Alpha terpineol	0.19	1186	11.593
14	Carvacrol methyl ether	0.12	1241	13.095
15	Thymol	0.31	1289	14.602
16	Carvacrol	86.91	1298	15.064
17	Carvacryl acetate	1.29	1370	17.2
18	Trans-caryophyllene	0.45	1417	18.676
19	Beta-bisabolene	1.05	1505	21.165
20	Cis-alpha-bisabolene	0.22	1506	22.057
Sum		99.37		

RI: Retention index / RT: Retention Time

3. 2. Microbial changes

In general, the total counts of mesophilic and psychrophilic bacteria in all four treatments increased over time(Fig 1 and Fig2). The growth rate of mesophilic bacteria in samples treated withSr-EO+SDS+citric acid and in samples treated by SDS+Sr-EO was lower than that in the other treatments. The International Commission on Microbiological Specifications for Foods (ICMSF, 1998) statedthat the acceptable level of total bacteria for fresh meat is <7log CFU/g and above this level, meatbegins to spoil and does not recommend.

The average mesophilic bacterial load increased in all control groups (Fig 1). It reached $7.24 \pm 0.16 \log$ (CFU/g) exceeding the acceptable level on day 6 of storage. However, for the other three treatments, the amount was less than seven until day 6 of storage. Given the obtained results and the determined minimum allowable microbial load for meat to be 7 logCFU/g, it was concluded that the population of mesophilic bacteria on day 6 of storage for the control group exceeded the allowable range; however, for other treatments, it exceeded the limit on day 9. In fact, the use of *Sr-EO*, SDS, and citric acid at the concentrations used could keep

the population of mesophilic bacteria within the permitted range until day 6 of storage.

The average psychrophilic bacteria load also increased for the control group and other treatment groups (Fig 2). The logarithm of psychrophilic bacterial load on day 6 of storage for the control, *Sr-EO* and *Sr-EO*+SDS+citric acid groups exceeded the acceptable value (7logCFU/g) however; for *Sr-EO*+SDS, it was less than 7 logs until day 6.

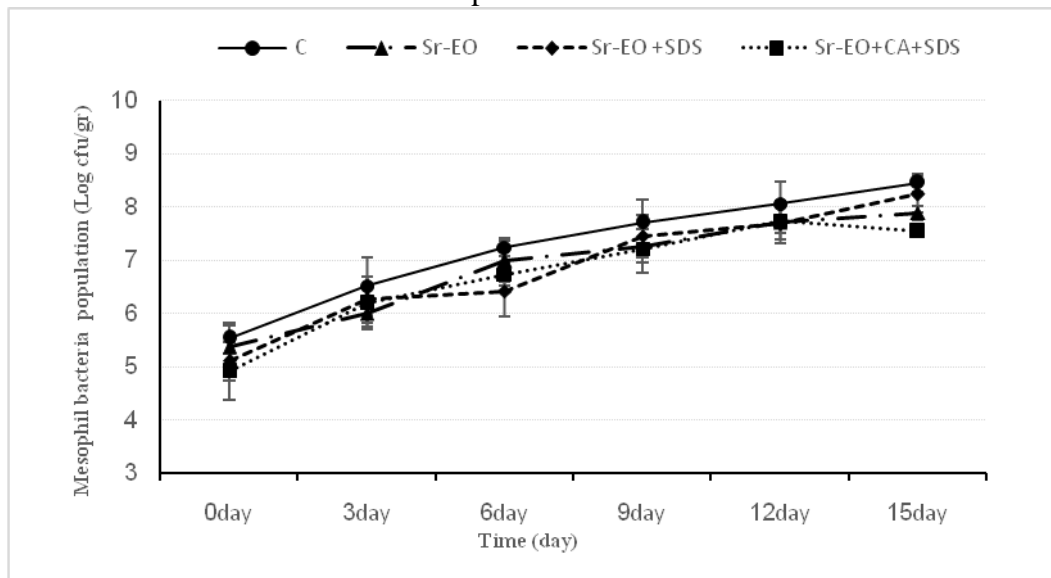


Fig. 1. Changes in total mesophilic count (log CFU/g) of chicken fillets during storage at 4 °C

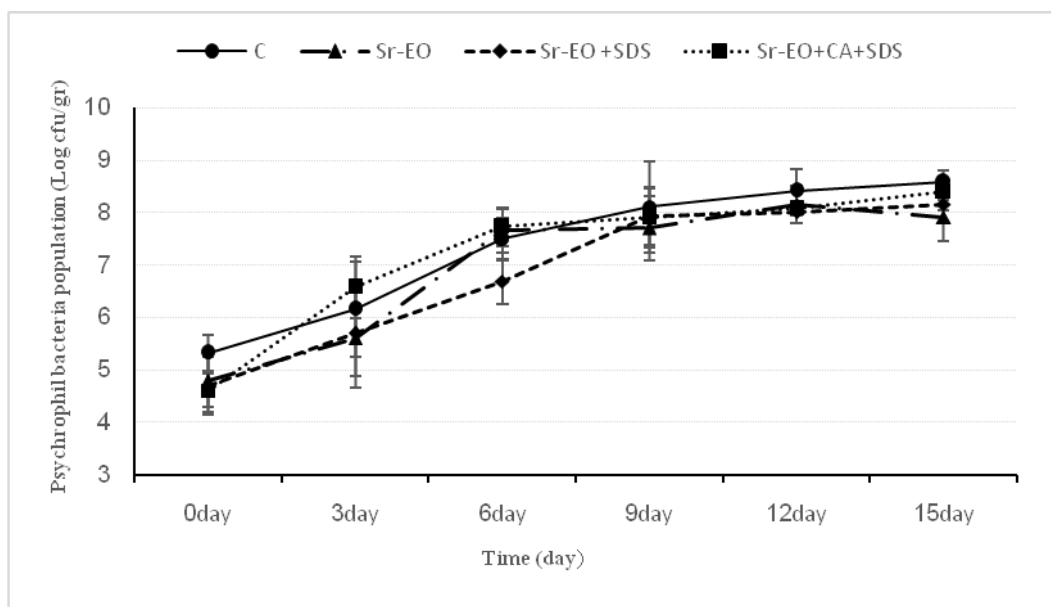


Fig. 2. Changes in total psychrophilic count (log CFU/g) of chicken fillets during storage at 4°C.

3. 3. Measurement of TBA value

According to Fig 3, the changes in the amount of thiobarbituric acid during the storage period of 15 days showed an increasing trend in all treatment groups. So that this increasing trend in the control group was similar to the *Sr*-EO+SDS+citric acid group. Also, *Sr*-EO and *Sr*-EO+SDS group showed similar changes.

At the end of the storage period, the highest amount of TBA in the *Sr*-EO+SDS+citric acid group was 0.60 ± 0.01 mg MDA/kg and the lowest amount of TBA in the *Sr*-EO group was 0.56 ± 0.6 mg MDA/kg and in the *Sr*-EO+SDS group was obtained as 0.56 ± 0.03 mg MDA/kg. Statistical analysis showed that the time has a significant effect on the amount of TBA ($P < 0.001$). But the group and the interaction between group and time have no significant effect ($P > 0.05$).

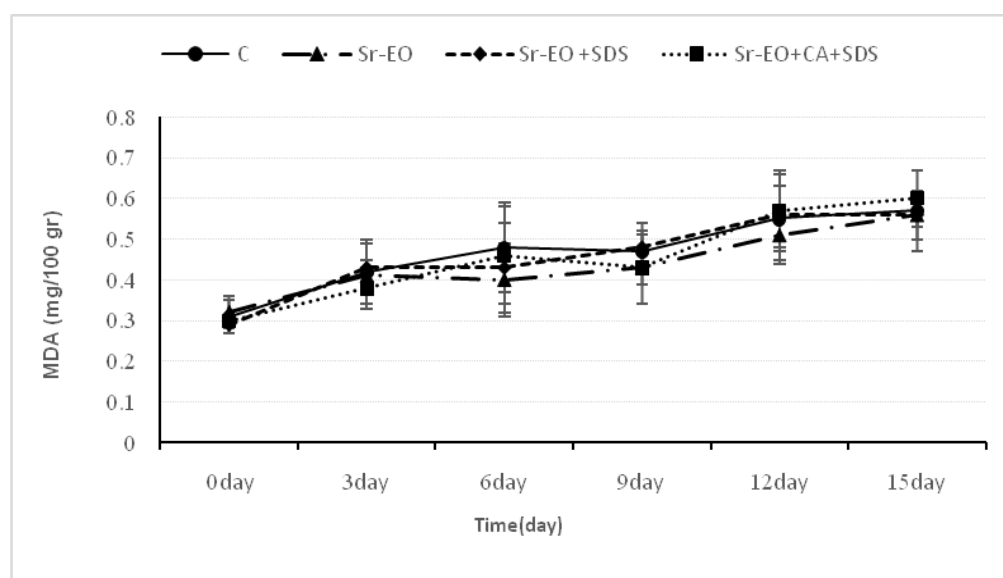


Fig. 3. Changes in TBA value (mg MDA/kg) of chicken fillets during storage at 4 °C.

3. 4. Measurement of TVN value

Figure 4 shows the changes of TVN in chicken fillet samples during the storage period. The trend of changes in volatile nitrogen bases was increasing for all treatments during 15 days of storage. This increase was not significant for control group, *Sr*-EO and *Sr*-EO+SDS+citric acid group. However for SDS+*Sr*-EO group, the increasing trend on day 0 was significantly different from other days. At the end of the storage period, the highest

TVN value (41.06 ± 8.29 g / 100 g) was observed for control sample and the lowest value (33 mg / 100 g) was found for *Sr*-EO+SDS+citric acid group followed by *Sr*-EO+SDS and *Sr*-EO groups. Given the allowable TVN values, it exceeded the permitted range in control group on day 12, however in other treatments, the rate of increasing trend for TVN was much slower than in control group and was in the desirable range until day 12.

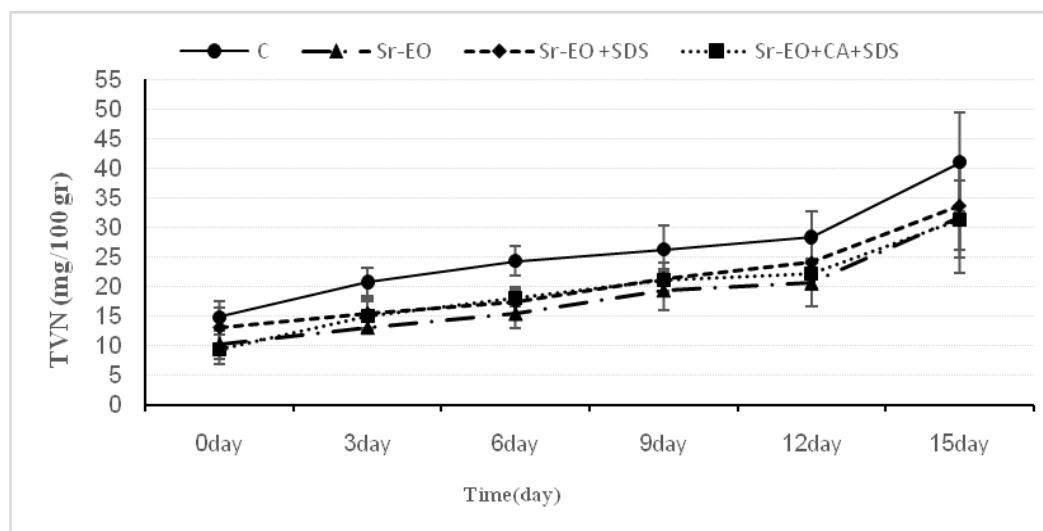


Fig. 4. Changes in TVN value (mg/100g) of chicken fillets during storage at 4 °c.

3. 5. Measurement of pH

The maximum and minimum pH values were 5.78 and 6.42 found respectively for the *Sr-EO* group and SDS+*Sr-EO* groups. In this study, the type of treatment did not have a significant effect on the pH value of

the samples. The pH of the treatment groups was almost the same and only in the *Sr-EO* treatment, the lowest pH values were observed on days 7, 9 and 15.

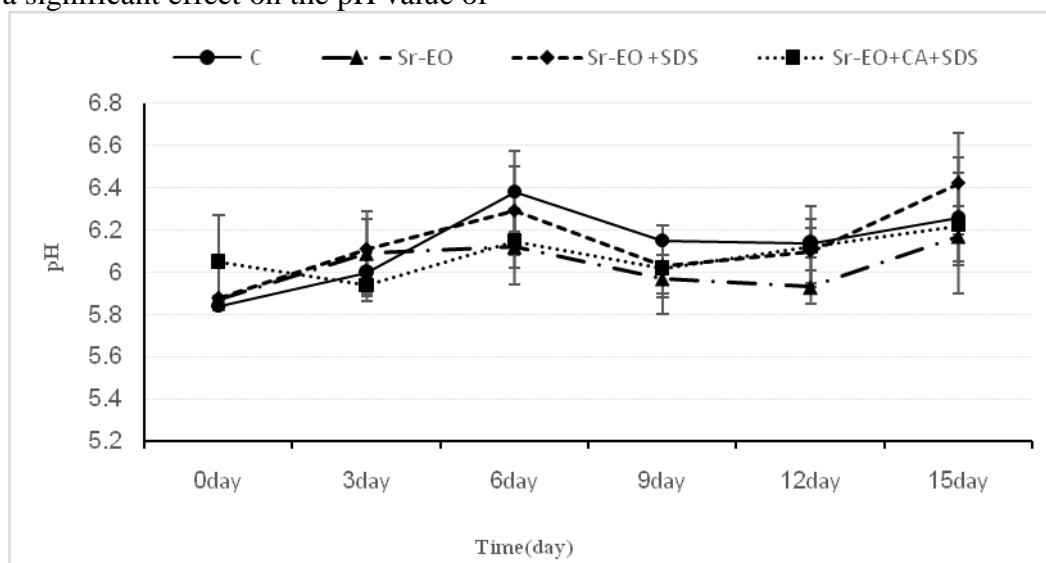


Fig. 5. Changes in pH value of chicken fillets during storage at 4 °c.

3. 6. Results of sensory tests

The results of the general acceptance of the samples during the storage period using the 5-point method are presented in Table 3. The statistical analysis between the groups by the type of treatment and time, with Friedman and Kruskal-Wallis test, showed that there is a significant

difference between the groups in terms of color, smell, texture and overall acceptance ($P < 0.05$).

The control group was unacceptable on day 6 regarding the examined four factors. The *Sr-EO* group was the best treatment, within the acceptable range until day 9 of storage. *Sr-EO+SDS* group was also in the

acceptable range until day 6 of storage in terms of color and texture, and day 9 of storage in terms of texture. The 4th treatment containing citric acid was not acceptable and had only an acceptable odor until day 15, while its color and texture were not acceptable. The data showed that citric acid, at the concentration used had limited antimicrobial effects on the samples. In this study, treatment with citric acid resulted undesirable color and texture changes. Therefore, *Sr*-EO+SDS +citric acid treatment was not effective in extending the shelf life of the raw chicken fillets. The best treatment in terms of sensory evaluation and panelist acceptance was the *Sr*-EO group until day 9 of storage, followed by *Sr*-EO+SDS group until day 6 of storage.

4. Discussion

The results showed that, in general, the treatment of chicken fillets with solutions containing *Sr*-EO with or without SDS was effective in reducing the bacterial load during storage in the refrigerator. As mentioned in results, *Sr*-EO+SDS could be more effective in terms of psychrophilic bacterial count, which might be due to the effect of SDS on growth inhibition.

SDS as an anionic surfactant able to disrupt membranes and denature proteins, so by the mechanism can kill bacteria [25]. (Woo et al., 2000). The decontamination of eggshell by SDS and citric acid has been already reported. Combination of SDS with components of essential oils may have a synergistic killing effect on bacteria [26]. (Moreira et al. (2007) reported that the effectiveness of essential oils in foods depends on the concentration of the EO used, the composition of the food and the

storage temperature because of the possible interactions between the EOs and the constituents of the food as well as the presence of protein and/or fat molecules that act as physical barriers against antimicrobial agents protecting the microorganisms against antimicrobial activity [27].

Chouliara et al. (2007) reported that chicken breast treated with oregano essential oil (1%) had a more potent protective effect than treated with 0.1% oregano essential oil. The concentration of 1% EO extended the shelf life up to 19-20 days while 0.1% EO increased the shelf life by only up to 1-2 days. The results obtained using *Sr*-EO and *Sr*-EO+SDS treatments in terms of increasing the shelf life of chicken fillets in the present study were consistent with the effect of the oregano EO in extending the shelf life up to 1-2 days [28]. The effect of *Saturejahortensis* essential oil on the quality and shelf life of chicken meat over refrigerated storage was also investigated [29]. In their study, pieces of meat were soaked for 2 hrs in *S. hortensis* treatment solutions at concentrations of 1, 3%, and 5%, and the results of microbial control showed that all three concentrations were effective in inhibiting the growth of microorganisms and increasing the shelf life of chicken meat by up to 2 days. A concentration of 5% had the greatest effect and reduced the total microbial load from 6.71 to 5.69 log cfu / g compared to the control. They used higher concentrations (up to 5%) and longer treatment times (2 h) in comparison to the current study, in which the treatment time of chicken fillets was 25 min and the concentration of *Sr*-EO was 0.5%.

Table.3. Changes sensory factors scores of chicken fillets during storage at 4°C

Group	Storage time (day)	Color	Odor	Texture	General acceptance
Control	0	Aa 5±0.00	Aa 5±0.00	Aa 5±0.00	Aa 5±0.00
Sr-EO		Aa 5±0.00	Aa 5±0.00	Aa 5±0.00	Aa 5±0.00
SDS+Sr-EO		Aa 5±0.00	Aa 5±0.00	Aa 5±0.00	Aa 5±0.00
Acid+SDS+Sr-EO		Aa 5±0.00	Aa 5±0.00	Aa 5±0.00	Aa 5±0.00
Control	3	ABa 4.33±0.44	Aab 4±0.00	ABab 3.66±0.33	Aab 3.66±0.33
Sr-EO		Aa 5±0.00	Aab 4.33±0.33	Aa 4.66±0.33	Aa 4.66±0.33
SDS+Sr-EO		ABab 4.33±0.33	Aa 4±0.00	Aab 4.66±0.33	Aab 4±0.00
Acid+SDS+Sr-EO		Bab 3.83±0.16	Aa 4±0.00	ABab 3.5±0.28	Aab 3.5±0.00
Control	6	Aa 3.8±0.61	Aab 3.33±0.33	Bab 2.60±0.30	Aab 2.83±0.44
Sr-EO		Aab 4.53±0.46	Aab 4.20±0.41	Aa 4.33±0.17	Aa 4.13±0.13
SDS+Sr-EO		Aab 4±0.23	Aa 3.86±0.46	ABab 4.06±0.29	Aab 3.86±0.24
Acid+SDS+Sr-EO		Aab 3.06±0.17	Aa 3.53±0.29	ABab 3.40±0.23	Aab 3.26±0.17
Control	9	Aa 3±1.15	Bb 1.86±0.86	Bbc 2.16±0.6	Bb 2.06±0.63
Sr-EO		Aab 4.33±0.33	Aab 4.86±0.13	Aa 4.33±0.33	Aa 4.2±0.2
SDS+Sr-EO		Aab 3.76±0.39	Aa 3.86±0.46	Aab 4±0.00	Aab 3.83±0.16
Acid+SDS+Sr-EO		Ab 2.80±0.11	Aa 3.53±0.29	ABab 3.46±0.29	ABab 3.16±0.16
Control	12	Bb 1±0.00	Bb 1.33±0.33	Bc 1.33±0.00	Bb 1±0.00
Sr-EO		Aab 3.83±0.44	Aa 3.56±0.29	Aa 4.16±0.16	Aa 3.83±0.44
SDS+Sr-EO		Aab 3.76±0.12	Aa 3.8±0.1	ABab 3.56±0.23	Aab 3.8±0.2
Acid+SDS+Sr-EO		ABb 2.73±0.26	Aa 3.6±0.16	ABab 2.83±0.16	Bb 1.90±0.45
Control	15	Ab 1±0.00	Bb 1±0.00	Ac 1±0.00	Ab 1±0.00
Sr-EO		Ab 2.83±0.44	ABb 2.90±0.45	Aa 2.86±0.46	Aa 2.86±0.46
SDS+Sr-EO		Aa 2.86±0.13	ABa 2.96±0.31	Ab 2.66±0.33	Ab 2.80±0.3
Acid+SDS+Sr-EO		Ab 2.43±0.43	Aa 3.66±0.16	Aab 2.20±0.2	Ab 1.50±0.28

- Different capital letters in each column indicate significant differences between treatments at each time point ($p < 0.05$).
- Different lowercase letters in each column indicate significant differences in each treatment at different times ($p < 0.05$).

Fat oxidation in meat generates compounds such as aldehydes, ketones, and alcohols that develop off-flavors and reduce the nutritional value of meat. The TBA value indicates fat oxidation, that is the amount of oxidation by-products, especially aldehydes, which are produced from the breakdown or oxidation of hydroperoxides. TBA is widely used to measure the degree of fat oxidation. Byun et al. (2003) reported that 2 mg of

malondialdehyde per kg of meat is an indicator of the onset of fat oxidation and sensory changes in chicken [30]. Teets and Were (2008) also reported that 3 mg malondialdehyde/kg indicated the onset of oxidative spoilage [31]. In the present study, the TBA value for all treatments was much lower than this range probably due to the low fat content of chicken fillet meat since the amount of fat and fatty acid composition are crucial factors fat

oxidation during storage. There was a significant difference among the treatments. On day 0, the TBA value for all treatments was approximately 0.3, which is in agreement with the results obtained by Hakim et al. [32]. In the current study, changes in TBA value during 15 days of refrigerated storage, except on day 9, increased for all treatments.

TVN value is an indicator of meat product freshness. It includes a wide range of volatile compounds such as ammonia, methylamine, dimethylamine, and other similar compounds that are produced when spoilage and microbial growth occur [33]. High levels of bacterial activity can increase the TVN value, because bacteria convert compounds such as tyramine oxide, peptides, and amino acids into volatile bases [34]. In a study conducted to preserve the quality of chicken breast fillets using sodium alginate incorporated with lemon verbena and clove essential oils, the lowest TVN value (36.66 mg/100 g) was obtained with 0.5% lemon verbena EO after 15 days of refrigerated storage, which is consistent with the results of the current study [35]. In a study by Ghanbari *et al.*, the effect of nanoemulsions of red grape seed essential oil at concentrations of 1%, 2%, and 5% on the shelf life of freshly packed chicken fillets at refrigerator temperature was investigated. The results showed that the nano essential oil of red grape seed at a concentration of 5% can reduce the increase in organic nitrogen bases and peroxide number during storage at 4°C by gradually releasing the substance [36]. Similar to the current study, other researchers observed that chicken meatball samples treated with thyme essential oil caused a significant decrease in TVN value compared to control samples during frozen storage periods. They reported that the addition of thyme essential oil had a positive effect on storage stability and a slight change in physical properties and quality attributes [37].

Changes in pH could be a useful way to evaluate qualitative changes in meat during storage. An increase in pH indicates a loss of quality. In contrast, the decrease in pH could be due to acid, which is a common metabolite from growth of a number of bacteria including lactic acid bacteria. The pH of the samples was not affected by various treatments in the current study, and the results are consistent with those obtained by Petrou *et al.* (2012), who investigated the natural antimicrobial effect of 1.5% chitosan, 0.25% v/w oregano, and their combination on the shelf life of chicken breast meat packed in modified atmosphere packaging during storage at 4°C [38]. These findings are consistent with the results obtained by Melo *et al.* (2012), who studied the effect of cellulose acetate-based active films at two concentrations of rosemary essential oil (20% and 50% v/w) on chicken breast meat during refrigerated storage. The initial average pH was 6.08 and the maximum value was 6.13 on day 6; therefore there was no significant difference in pH values among the treatments [39].

Changes in sensory properties are one of the main reasons for the reduced shelf life of meat. Microbial growth and fat oxidation result in the production of undesirable metabolites, thereby reducing sensory properties [22]. The results of the sensory evaluation are consistent with those obtained by Hartanti *et al.* (2019), who investigated the effects of lemongrass and lemon basil essential oils individually or in combination at different concentrations on microbial quality and physical properties, including color, odor, texture, glaze, and slime of chicken fillet meat during refrigerated storage for 9 days. The results showed that the combined treatment had a better performance and was acceptable in terms of color and glaze until day 9 of storage. In addition, the rate of change in texture and odor in the combined treatment was slower than that in the EO and control groups [40]. Heydarian *et al.* (2015) also

studied the antioxidant and antimicrobial effects of aqueous rosemary extract at concentrations of 1, 3% and 5% on the quality and shelf life of chicken meat for 5 d at 4 °C. Sensory evaluation results revealed that the sensory scores decreased significantly over time. The 3% treatment resulted in the highest sensory scores compared to the control and the 1 and 5% treatments. The 1 and 5% treatments were acceptable until day 3 and the 3% treatment until day 5[41].

5. Conclusion

Given the sensory results and total acceptance of chemical and bacterial properties, the shelf life of chicken fillets for the control group was 2- 3 days, for Sr-EO group 3- 4 days, for Sr-EO+SDS group 6-7 days, and for Sr-EO+SDS+citric acid group 3-4 days. The results showed that SDS+Sr-EO treatment was more effective in extending the shelf life of chicken fillets; therefore, it can be used to increase the refrigerated storage time of chicken meat by up to 4 days or to manufacture new products with a special aroma to meet consumer expectations. Further research is required to determine the best combination to achieve better results.

6. Acknowledgments

We would like to express our appreciation of the Research Council of Shahid Chamran University of Ahvaz, Iran, for their financial support.

7. References

- [1]Jimenez, S. M., Salsi, M. S., Tiburzi, M. C., Rafaghelli, R. C., &Pirovani, M. E. (1999). Combined use of acetic acid treatment and modified atmosphere packaging for extending the shelf-life of chilled chicken breast portions. *Journal of Applied Microbiology*, 87(3), 339-344.
- [2]Azlin-Hasim, S., Cruz-Romero, M. C., Morris, M. A., Cummins, E., & Kerry, J. P. (2015). Effects of a combination of

antimicrobial silver low density polyethylene nanocomposite films and modified atmosphere packaging on the shelf life of chicken breast fillets.*Food Packaging and Shelf Life*, 4, 26-35.

[3]Fратиanni, F., De Martino, L., Melone, A., De Feo, V., Coppola, R., &Nazzaro, F. (2010). Preservation of chicken breast meat treated with thyme and balm essential oils. *Journal of food science*, 75(8), 528-535.

[4]Jafarzadeh, k. K.; Aghazadeh, M. M.; Sharifan, A. and Larijani, K. (2010). Investigation of effect of the rosemary essential oil on growth of staphylococcus aureus in commercial instant soup.*Iranian comparative pathobiology*, 7(2), 255-263

[5] Jamzad, Z. (1996). Saturejarechingeri (Labiatae)—a new species from Iran.*Annalen des Naturhistorischen Museums in Wien.Serie B fürBotanik und Zoologie*, 75-77.

[6] Nooshkam, A., MajnounHoseini, N., Hadian, J., Jahansooz, M. R., Khavazi, K., Salehnia, A., &Hedayatpour, S. (2016). The Effect of Irrigation Intervals on Quantitative and Qualitative Yields of Two Savory Species (Saturejakhuzestanica and S. rechingeri). *Journal of Plant Productions (Agronomy, Breeding and Horticulture)*, 39(1), 11-20.

[7] Hadian, J., Akramian, M., Heydari, H., Mumivand, H., &Asghari, B. (2012). Composition and in vitro antibacterial activity of essential oils from four Satureja species growing in Iran.*Natural product research*, 26(2), 98-108.

[8]Rezvanfar, M. A., Sadrkhanlou, R. A., Ahmadi, A., Shojaei-Sadee, H., Rezvanfar, M. A., Mohammadirad, A., ... &Abdollahi, M. (2008). Protection of cyclophosphamide-induced toxicity in reproductive tract histology, sperm characteristics, and DNA damage by an herbal source; evidence for role of free-radical toxic stress.*Human & experimental toxicology*, 27(12), 901-910.

[9]Morales-delaNuez, A., Moreno-Indias, I., Sánchez-Macías, D., Capote, J., Juste,

- M. C., Castro, N., ...&Argüello, A. (2011). Sodium dodecyl sulfate reduces bacterial contamination in goat colostrum without negative effects on immune passive transfer in goat kids. *Journal of dairy science*, 94(1), 410-415.
- [10]Stelzleni, A. M., Ponrajan, A., & Harrison, M. A. (2013). Effects of buffered vinegar and sodium dodecyl sulfate plus levulinic acid on Salmonella Typhimurium survival, shelf-life, and sensory characteristics of ground beef patties. *Meat science*, 95(1), 1-7.
- [11] Li, Y., & Wu, C. (2013). Enhanced inactivation of Salmonella Typhimurium from blueberries by combinations of sodium dodecyl sulfate with organic acids or hydrogen peroxide. *Food Research International*, 54(2), 1553-1559.
- [12] Lu, Y., & Wu, C. (2012). Reductions of Salmonella enterica on chicken breast by thymol, acetic acid, sodium dodecyl sulfate or hydrogen peroxide combinations as compared to chlorine wash. *International journal of food microbiology*, 152(1-2), 31-34.
- [13] Park, Y. B., Guo, J. Y., Rahman, S. M. E., Ahn, J., & Oh, D. H. (2009). Synergistic effect of electrolyzed water and citric acid against Bacillus cereus cells and spores on cereal grains. *Journal of food science*, 74(4), M185-M189.
- [14] Aynewa, D., Gizaw, Z., & Haile, A. F. (2021). Assessment of bacteriological quality of sheep carcasses, effect level of 2.5% citric acid spray on bacterial contamination of meat, and hygiene practices of workers in a selected abattoir in Debrezeit town, central Ethiopia. *Environmental Health Insights*, 15, 11786302211037555.
- [15]Noori, S., Zeynali, F., &Almasi, H. (2018). Antimicrobial and antioxidant efficiency of nanoemulsion-based edible coating containing ginger (Zingiberofficinale) essential oil and its effect on safety and quality attributes of chicken breast fillets. *Food control*, 84, 312-320.
- [16]Zamuz, S., López-Pedrouso, M., Barba, F. J., Lorenzo, J. M., Domínguez, H., & Franco, D. (2018). Application of hull, bur and leaf chestnut extracts on the shelf-life of beef patties stored under MAP: Evaluation of their impact on physicochemical properties, lipid oxidation, antioxidant, and antimicrobial potential. *Food Research International*, 112, 263-273.
- [17]Anonymous (1386). Microbiology of food and animal reference method for counting the total count of microorganisms in 30 degrees Celsius, *Iranian National Standard no.5272*, the first revision.
- [18] Xiong, Z., Sun, D. W., Pu, H., Xie, A., Han, Z., & Luo, M. (2015). Non-destructive prediction of thiobarbituric acid reactive substances (TBARS) value for freshness evaluation of chicken meat using hyperspectral imaging. *Food chemistry*, 179, 175-181.
- [19] Malle, P., &Poumeyrol, M. (1989). A new chemical criterion for the quality control of fish: trimethylamine/total volatile basic nitrogen (%). *Journal of food protection*, 52(6), 419-423.
- [20]. Parvaneh, Vida (1377). Food quality control and chemical testing, Tehran university press, fourth edition, pages: 251-249.
- [21] Fan, W., Sun, J., Chen, Y., Qiu, J., Zhang, Y., & Chi, Y. (2009). Effects of chitosan coating on quality and shelf life of silver carp during frozen storage. *Food chemistry*, 115(1), 66-70.
- [22] Hassanzadeh, P., Tajik, H., Rohani, S. M. R., Moradi, M., Hashemi, M., &Aliakbarlu, J. (2017). Effect of functional chitosan coating and gamma irradiation on the shelf-life of chicken meat during refrigerated storage. *Radiation physics and chemistry*, 141, 103-109.
- [23]Ojagh, S. M., Rezaei, M., Razavi, S. H., & Hosseini, S. M. H. (2010). Effect of chitosan coatings enriched with cinnamon oil on the quality of refrigerated rainbow trout. *Food chemistry*, 120(1), 193-198.
- [24] Raeisi, M., Tabaraei, A., Hashemi, M., &Behnampour, N. (2016). Effect of

sodium alginate coating incorporated with nisin, Cinnamomumzeylanicum, and rosemary essential oils on microbial quality of chicken meat and fate of *Listeria monocytogenes* during refrigeration. *International journal of food microbiology*, 238, 139-145.

[25] Woo I., Rhee I. and Park H. (2000). Differential damage in bacterial cells by microwave adiation on the basis of cell wall structure. *Applied. Environ. Microbiology*. 66: 2243-2247.

[26] Maktabi, S., Zarei, M., & Rashnavady, R. (2018). Effect of sequential treatments with sodium dodecyl sulfate and citric acid or hydrogen peroxide on the reduction of some foodborne pathogens on eggshell. *Iranian journal of veterinary research*, 19(2), 113.

[27] Moreira, M. R., Ponce, A. G., Del Valle, C. E., & Roura, S. I. (2007). Effects of clove and tea tree oils on *Escherichia coli* O157: H7 in blanched spinach and minced cooked beef. *Journal of Food Processing and Preservation*, 31(4), 379-391.

[28] Chouliara, E., Karatapanis, A., Savvaidis, I. N., & Kontominas, M. G. (2007). Combined effect of oregano essential oil and modified atmosphere packaging on shelf-life extension of fresh chicken breast meat, stored at 4 C. *Food microbiology*, 24(6), 607-617.

[29] Nusrat Elahi, K., Barzegar, H., Joyandeh, H. and Ghorbani, M. (2018). Investigation of the effect of savory extract (*Saturejahortensis*) on the quality and shelf life of refrigerated chicken. *Iranian Food Science and Technology*, 15 (82), 167-176.

[30] Byun, J. S., Min, J. S., Kim, I. S., Kim, J. W., Chung, M. S., & Lee, M. (2003). Comparison of indicators of microbial quality of meat during aerobic cold storage. *Journal of food protection*, 66(9), 1733-1737.

[31] Teets, A. S., & Were, L. M. (2008). Inhibition of lipid oxidation in refrigerated and frozen salted raw minced chicken breasts with electron beam irradiated

almond skin powder. *Meat Science*, 80(4), 1326-1332.

[32] Hakim, H., Fazlara, A. and Tadaini, M. (1397). The effect of chitosan coating containing oregano essential oil on the shelf life of chicken fillet meat during storage at refrigerator temperature. *Iranian Food Science and Technology*, 15 (75), 45-35.

[33] Raeisi, M., Tabaraei, A., Hashemi, M., & Behnampour, N. (2016). Effect of sodium alginate coating incorporated with nisin, Cinnamomumzeylanicum, and rosemary essential oils on microbial quality of chicken meat and fate of *Listeria monocytogenes* during refrigeration. *International journal of food microbiology*, 238, 139-145.

[34] López-Caballero, M. E., Gómez-Guillén, M. C., Pérez-Mateos, M., & Montero, P. (2005). A chitosan–gelatin blend as a coating for fish patties. *Food hydrocolloids*, 19(2), 303-311

[35] Hosseini, M., Jamshidi, A., Raeisi, M., & Azizzadeh, M. (2021). Effect of sodium alginate coating containing clove (*Syzygium Aromaticum*) and lemon verbena (*Aloysia Citriodora*) essential oils and different packaging treatments on shelf life extension of refrigerated chicken breast. *Journal of Food Processing and Preservation*, 45(3), e14946.

[36] Ghanbari, M., Motallebi, A., Rokni, N., & Anvar, A. (2020). Evaluation of red grape seed essential oil nanoemulsion (*Vitisvinefera*) on the shelf life of fresh packaged chicken fillets during refrigerated storage at 4° C. *Arch. Pharm. Pract.*, 1, 120.

[37] Sorour, M. A., El-Hamied, A., Adel, A., Mohamoud, E. S. A., & Mahmoud, A. R. (2022). Anti-Oxidative Impact of Liquid Smoke and Thyme Essential Oil on the Quality Characteristics of Chicken and Turkey Meatballs Products during Frozen Storage. *European Journal of Nutrition & Food Safety*, 14(3), 14-26.

[38] Petrou, S., Tsiraki, M., Giatrakou, V., & Savvaidis, I. N. (2012). Chitosan dipping or oregano oil treatments, singly or

combined on modified atmosphere packaged chicken breast meat. *International journal of food microbiology*, 156(3), 264-271.

[39] Melo, A. A. M. D., Geraldine, R. M., Silveira, M. F. A., Torres, M. C. L., Fernandes, T. H., & Oliveira, A. N. D. (2012). Microbiological quality and other characteristics of refrigerated chicken meat in contact with cellulose acetate-based film incorporated with rosemary essential oil. *Brazilian Journal of Microbiology*, 43, 1419-1427.

[40] Hartanti, D., Nuritasari, A., & Hamad, A. (2019). A combination of lemongrass and lemon basil essential oils inhibited bacterial growth and improved the shelf life of chicken fillets. *Asia-Pacific Journal of Science and Technology*, 24(04), 1-9.

[41] Heydarian, M. T., JebelliJavan, A., & Jekar, M. (2015). Antimicrobial and antioxidant effects of rosemary extract on quality and shelf life of raw chicken during refrigerated storage. *Research and Innovation in Food Science and Technology*, 4(2), 131-142.



بررسی خواص میکروبی و شیمیایی فیله مرغ تیمار شده با اسانس مرزه رشینگری، سدیم دودسیل سولفات و اسید سیتریک در طی نگهداری در یخچال

سیاوش مکتبی^{۱*}، زینب صفدری^۲، مهدی پورمهدی بروجنی^۳

۱- دانشیار گروه بهداشت مواد غذایی، دانشکده دامپزشکی، دانشگاه شهید چمران اهواز

۲- فارغ التحصیل کارشناسی ارشد بهداشت و کنترل کیفی مواد غذایی، دانشکده دامپزشکی، دانشگاه شهید چمران اهواز

۳- استاد گروه بهداشت مواد غذایی، دانشکده دامپزشکی، دانشگاه شهید چمران اهواز

اطلاعات مقاله

چکیده

تاریخ های مقاله :

تاریخ دریافت: ۱۴۰۱/۷/۱۹

تاریخ پذیرش: ۱۴۰۱/۱۲/۱

کلمات کلیدی:

گوشت مرغ،

فیله مرغ،

اسانس مرزه رشینگری،

سدیم دودسیل سولفات،

اسیدسیتریک،

مدت ماندگاری

DOI: 10.22034/FSCT.20.140.1

DOR: 20.1001.1.20088787.1402.20.140.1

* مسئول مکاتبات:

s.maktabi@scu.ac.ir

در این پژوهش، تاثیر اسانس گیاه مرزه رشینگری (*Saturejarechingeri*)، سدیم دودسیل سولفات (SDS) و اسیدسیتریک بر روی افزایش ماندگاری فیله مرغ در یخچال بررسی گردید. طی ۳ تکرار و هر بار ۲۸ قطعه فیله مرغ تهیه و به ۴ گروه تقسیم شدند. هر گروه به مدت ۱۵ دقیقه در محلول های حاوی سرم فیزیولوژی (کنترل)، ۰/۵ درصد اسانس مرزه رشینگری، ۰/۵ درصد اسانس SDS، ۰/۵ درصد اسانس SDS+اسیدسیتریک (۰/۱) قرار گرفته و سپس به مدت ۱۵ روز در یخچال نگهداری شدند. آزمون های شمارش باکتریایی (مزوفیل و سایکروفیل)، آزمون های شیمیایی (TVN, TBA, pH) و حسی طی روزهای ۰، ۳، ۶، ۹، ۱۲ و ۱۵ بر روی نمونه ها انجام گرفت. نتایج نشان داد بار باکتریایی مزوفیل و سایکروفیل با گذر زمان در هر چهار گروه روند افزایشی دارد، اما تعداد باکتری ها در گروه اسانس SDS+اسیدسیتریک، برای شاخص میکروبی مزوفیل و در گروه اسانس SDS، برای شاخص میکروبی ساکروفیل کمتر از گروه های دیگر بود. در پایان مدت زمان نگهداری، بیشترین میزان TVN مربوط به گروه کنترل (۸/۲۹ ± ۴۱/۰۶ گرم در ۱۰۰ گرم) و کمترین میزان در محدوده ۳۳ میلی گرم در ۱۰۰ گرم، به ترتیب مربوط به گروه اسانس SDS+اسیدسیتریک و گروه اسانس SDS و گروه اسانس بود. در خصوص فاکتور TBA و pH اختلاف قابل توجهی بین گروه های تیماری مشاهده نشد. از نظر فاکتورهای حسی نیز، گروه کنترل در روز ششم نگهداری در محدوده نامطلوب و غیرقابل پذیرش قرار گرفت. درحالی که گروه اسانس و گروه اسانس SDS به ترتیب تا روز نهم و ششم نگهداری در محدوده مطلوب پذیرش انسانی قرار داشتند. نتایج نشان داد که استفاده از تیمار اسانس SDS در افزایش مدت زمان نگهداری فیله مرغ موثرتر عمل نموده و قادر است که زمان نگهداری گوشت مرغ را در یخچال تا ۴ روز افزایش دهد. این روش می تواند به عنوان روشی سودمند جهت افزایش زمان نگهداری گوشت مرغ در شرایط یخچالی به کار رود.